

Original Article

Rapid Production of *Candida albicans* Chlamydo spores in Liquid Media under Various Incubation Conditions

Zavalza-Stiker Alicia¹, Ortiz-Saldivar Blanca¹, García-Hernández Mariana¹,
Castillo-Casanova Magdalena¹, Bonifaz Alexandro²

¹Mycology Laboratory of Chemistry School, Universidad Nacional Autónoma de San Luis Potosí, México

²Mycology Department, Hospital General de México, OD. México D.F.

Dr. Balmis 148, Col Doctores, México DF.

Abstract

The production of chlamydo spores is a diagnostic tool used to identify *Candida albicans*; these structures also represent a model for morphogenetic research. The time required to produce them with standard methods is 48-72 hours in rice meal agar and tensoactive agents. This time can be shorted using liquid media such as cornmeal broth (CMB) and dairy supplements.

Five media were tested: CMB plus 1% Tween-80, CMB plus 5% milk, CMB plus 5% milk serum, milk serum, and milk serum plus 1% Tween-80, under different incubation conditions: at 28°C and 37°C in a metabolic bath stirring at 150rpm, and at 28°C in a culture stove. The reading time points were established at 8 and 16 hours. The best results were obtained at 16 hours with CMB plus 5% milk under incubation at 28°C and stirring at 150 rpm. The next most efficient methods were CMB plus 5% milk serum and CMB plus 1% Tween-80, under the same incubation conditions. The other media were ineffective in producing chlamydo spores. The absence of stirring at 28°C prevented the formation of chlamydo spores within the set time points, and incubation at 37°C decreased their production.

This paper reports that the time to form *C. albicans* chlamydo spores can be reduced.

Key words: *Candida albicans*, chlamydo spores, liquid media, cornmeal

Introduction

The formation of chlamydo spores is a morphologic feature of *Candida albicans* widely used in the laboratory for identification purposes. Together with the serum filamentation test, it constitutes a straightforward and low-cost etiologic diagnostic tool^{1, 2}. Another important aspect of the production and purification of *Candida albicans* chlamydo spores is the recent interest raised by the isolation of these structures in morphogenetic studies³. Traditionally, chlamydo spores have been obtained using a standard medium, i.e., cornmeal agar (CMA) with added Tween-80, in a minimum period of 48-72 hours². Some authors have described changes in this medium aimed at more efficient production; adding milk has resulted in an increased proportion of chlamydo spores

from 21.4% to 95.5% in 48 hours⁴. Another variant reported is the use of skim milk and the addition of solid media from processed cheeses, which led to an abundant and quicker development of chlamydo spores than with the standard method⁵. With cornmeal broth (CMB), in the same medium, with 5% milk or 1% veal serum, it has been possible to produce chlamydo spores in periods ranging from 20 to 72 hours, at an optimal temperature within a range of 24°C to 30°C⁶. This study intends to define conditions providing an abundant production of chlamydo spores in a shorter period than the times reported in the literature.

Materials and methods

Five liquid media were used: CMB plus 1% Tween-80, CMB plus 5% milk, CMB plus 5% milk serum, milk serum, and milk serum plus 1% Tween-80. The CMB was prepared at a 3% concentration in distilled water. Milk serum was

Correspondence: Dr. Alexandro Bonifaz

Sanchez Azcona 317 int 202, Col. Del Valle, CP, 03020
México D.F.

obtained from non-pasteurized cow's milk, with enzymes for coagulation of casein and separation by means of filtration. It was preserved frozen at -20°C . Pasteurized milk was used in the medium consisting of CMB with milk.

Candida species, isolated from different patient specimens and preserved in the strain collection at the Mycology Laboratory, were used in this study; negative controls were one species of *C. guilliermondii* and one of *C. parapsilosis*. Yeasts of the study were isolated and classified by traditional routine: isolation in three media: Biggy-Nickerson, CHROMagar-*Candida* and Sabouraud agar. Each of the strains has a precise typing scheme consisting of morphologic, physiologic as serum-filamentation; pseudohyphae and chlamydospore production in cornmeal + Tween 80 1% (agar-plate) and biochemical tests of carbohydrate assimilation with API 20C. Pre-culture used was potato dextrose agar (PDA), with two days of yeast growth. Broth media were inoculated with $100\ \mu\text{l}$ of turbidity-adjusted suspensions at an approximate concentration of 1×10^6 .

The tests were performed in a tube and incubated in a metabolic bath at 28°C and 37°C , stirred at 150 rpm, and in a tube with stove incubation at 28°C (without stirring). Results were read at 8 and 16 hours.

Chlamydospores were observed by a direct examination. A drop of inoculated broth media was placed (after vortex-agitation) between a microscope-slide and a cover-slide; chlamydospore production was quantitated by a semi-quantitative

method: we counted a number of them by microscopic fields and quantification was made by two observers.

Results

Results are reported in Tables 1 and 2. Table 1 indicates a percent of *C. albicans* strains with formation of chlamydospores in different liquid media under incubation and metabolic bath at 28°C and stirring at 150 rpm. Table 2 reports a semi-quantitative assessment of *C. albicans* chlamydospore production at 16 hours in different liquid media, with metabolic bath incubation at 28°C and 37°C and stirring at 150 rpm. (Fig. 1 and Fig. 2)

Discussion

Chlamydospores of *Candida albicans* depends on a regulatory protein which influences the transition from yeast to hyphae; 6 genes are needed that depend on certain conditions for their activation such as oxygen, pH and several substrates^{7, 8}.

In our study, we obtained good results, particularly in three of the culture media: CMB plus 5% milk, CMB plus 5% milk serum and CMB plus Tween-80. In the other two media, milk serum and milk serum plus Tween 80, no chlamydospores were formed under the established conditions. The production of chlamydospores was most effective and was achieved at 16 hours in the medium of CMB plus 5% milk. (Table 1)

Table 1. Percent of formation of chlamydospores in different liquid media

Strain	No. strains	Culture media					
		CMB + 1% of Tween 80		CMB + 5% of milk		CMB + 5% of milk serum	
		8 h (%)	16 h (%)	8 h (%)	16 h (%)	8 h (%)	16 h (%)
<i>C. albicans</i>	25	0	66	16.6	100	16.6	100
<i>C. guilliermondii</i>	1	0	0	0	0	0	0
<i>C. parapsilosis</i>	1	0	0	0	0	0	0

CMB= Cornmeal broth. Incubation: Metabolic bath at 28°C and stirring 150 rpm.

Table 2. Production of chlamydospores in semi-quantitative assessment of *C. albicans* at 16 hours in different liquid media

Strain	No. strains	Culture media					
		CMB + 1% of Tween 80		CMB + 5% of milk		CMB + 5% of milk serum	
		28°C	37°C	28°C	37°C	28°C	37°C
<i>C. albicans</i>	25	++	+	+++	+	++	+
<i>C. guilliermondii</i>	1	0	0	0	0	0	0
<i>C. parapsilosis</i>	1	0	0	0	0	0	0

CMB= Cornmeal broth. Incubation: Metabolic bath at 28°C and stirring 150 rpm.

Chlamydospore production: + = limited, ++ = moderate, +++ = abundant.

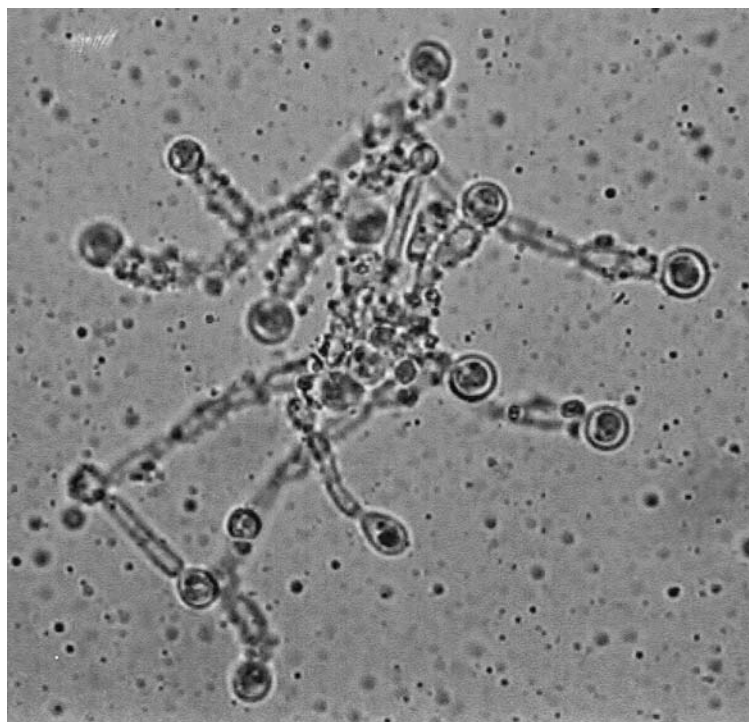


Fig. 1. Chlamydospores of *C. albicans* from CMB plus 5% milk media. (Direct exam, 10 $\times$)

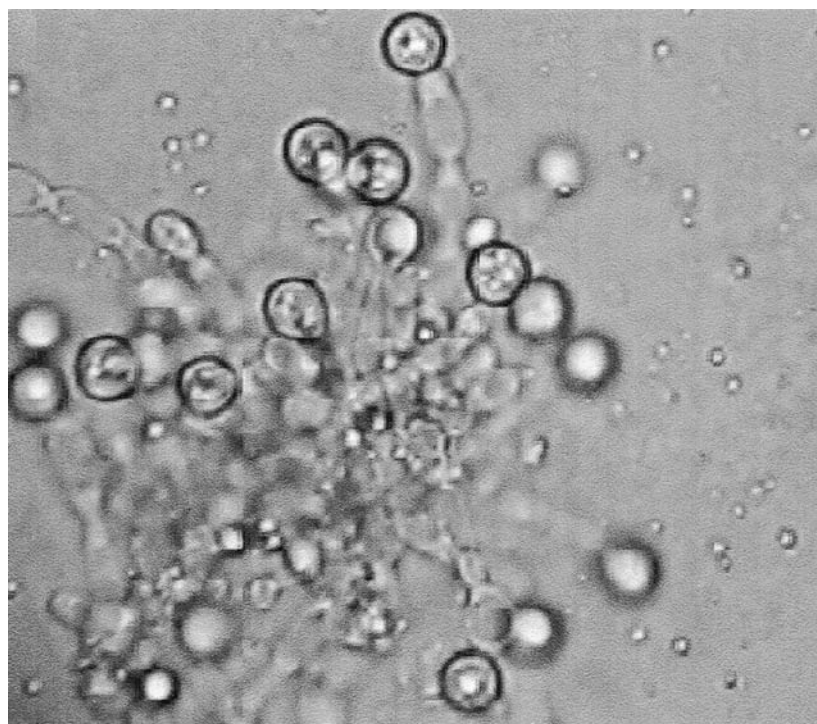


Fig. 2. Chlamydospores of *C. albicans* from CMB plus 5% milk media. (Direct exam, 40 $\times$)

The comparison of these results with the literature reports confirmed the effectiveness of milk as a factor that helps in the formation of *C. albicans* chlamydospores, in combination with CMB. To a great extent, milk was also found to favor the production of mycelia from the early stages (8 hours). As regards the time necessary

for the production of chlamydospores, stirring in a metabolic bath resulted in a 20% reduction in the time reported by Nakamoto⁶⁾, which was 20 hours under similar conditions (without stirring). A shorter time to produce these structures represents an advantage in the daily laboratory diagnostic work. Likewise,

morphogenetic research would benefit from a more effective separation of chlamydospores due to the possibility of producing large amounts under the above established conditions, besides the reduced time.

Casein-free milk (milk serum) was also found to favor the production of chlamydospores, with greater effectiveness than Tween 80 but less than whole milk, and with the added advantage that it can be preserved frozen for long periods of time.

In conclusion, it is important to reduce the time for chlamydospore formation. We also emphasize that the production took place in media without casein, proving that this protein is not necessary for chlamydospore-production

References

- 1) Odds FC: *Candida* and Candidosis. 2nd edition. Bailliere Tindall, London, 1988.
- 2) Kwon Chung KJ, Bennett JE: Candidiasis. *In*: Medical Mycology (Lea & Febiger ed), Chap. 13, pp.280-336, Philadelphia, 1992.
- 3) Fabry W, Schmid EN, Schrap M, Ansorg R: Isolation and purification of chlamydospores of *Candida albicans*. *Med Mycol* **41**: 53-58, 2003.
- 4) Jitsuron S, Kiamsiri S, Pattararangrong N: New milk medium for germ tube and chlamydoconidia production by *Candida albicans*. *Mycopathologia* **123**: 95-98, 1993.
- 5) Viddoto V, Caramello S, Gallo MG: A new medium for the production of chlamydoconidia by *Candida albicans*. *Mycopathologia* **95**: 73-75 1986.
- 6) Nakamoto S: Promotion of chlamydoconidium formation in *Candida albicans* by corn meal broth incubation. *Med Mycol* **36**: 123-125, 1998.
- 7) Sonneborn A, Bockmuhl DP, Ernst JF: Chlamydospore formation in *Candida albicans* requires the Efg1p morphogenetic regulator. *Infect Immun* **67**: 5514-5517, 1999.
- 8) Nobile CJ, Bruno VM, Richard ML, Davis DA, Mitchell AP: Genetic control of chlamydospore formation in *Candida albicans*. *Microbiology* **149**: 3629-3637, 2003.
- 9) Al-Hedaithy SS, Fotedar R: Recovery and studies on chlamydospore-negative *Candida albicans* isolated from clinical specimens. *Med Mycol* **40**: 301-306, 2002.
- 10) Sullivan D, Coleman D: *Candida dubliniensis*: characteristics and identification. *J Clin Microbiol* **36**: 329-334, 1998.