

Short Report

Evaluation of a Newly Developed Identification Kit, RID Zyme CAS Test, for *Candida albicans*

Reiko Tanaka, Junko Ito, Ayaka Sato, Kazuko Nishimura

Research Center for Pathogenic Fungi and Microbial Toxicoses, Chiba University
1-8-1 Inohana, Chuo-ku, Chiba, 260-8673 Japan

[Received: 26, October 2004. Accepted: 28, December 2004]

Abstract

To evaluate a newly developed identification kit, the RID Zyme CAS test for *Candida albicans*, 1136 *C. albicans* and 403 non-*albicans* *Candida* strains were tested. Distinction of medically important non-*albicans* strains, with the exception of *C. dubliniensis*, was obtained. These results show that this new kit is simple and effective for the identification of *C. albicans* in clinical samples. Furthermore, the one hour period for identification makes it very attractive.

Key words: *Candida albicans*, newly developed identification kit, RID Zyme CAS test

Candida albicans is the most frequently isolated yeast pathogen, and candidiasis is increasingly a complication in both immunocompromised and immunocompetent individuals. Immediate identification of the pathogen is usually required, however, conventional methods such as the API 20C AUX and ID32C systems (bioMerieux Japan, Tokyo, Japan) take more than 24 to 48 hours to give results. Several researchers have attempted to develop rapid methods for the identification of *C. albicans*. Perry *et al.*¹⁾ and Dalton *et al.*²⁾ reported rapid methods that make use of the fluorogenic substrate (4-methylumbelliferyl-*N*-acetyl- β -D-galactosaminide) of β -galactosaminidase (EC 3.2.1.30). Perry *et al.*¹⁾ reported a modification of their previous method, which involves alteration of the test substrate and inclusion of a second substrate in one reaction tube and provides a colorimetric rather than a fluorometric reaction product³⁾.

Here, we report the evaluation of a newly developed identification kit, the RID Zyme CAS test, on 1136 *C. albicans* and 403 non-*albicans* *Candida* species and five serotypes of *Cryptococcus neoformans*.

Strains used in this study are listed in Tables 1 and 2. These strains are registered at the Research Center for Pathogenic Fungi and Microbial Toxicoses of Chiba University and

were cultured on potato dextrose agar (PDA; Difco Laboratories, Detroit, MI, USA) at 25°C for 48 hours. Most of the strains were previously identified using the ID32C system (bioMerieux Japan) and/or the Candida Check system (Mitsubishi Kagaku Iatron, Inc., Tokyo, Japan), and were genotyped by PCR of 25S rDNA⁴⁾ and the topoisomerase II gene⁵⁾. Some strains were purchased from ATCC: American Type Culture Collection (Manassas, VA, USA), CBS: Centraalbureau voor Schimmelcultures, (Delft, the Netherlands), IFO: Institute for Fermentation, Osaka (Osaka, Japan) or NBRC: National Institute of Technology and Evaluation Biological Resource Center (Chiba, Japan) as references.

The RID Zyme CAS test (Mitsubishi Kagaku Iatron, Inc.) was put out to the market in 2004. Although two of the enzymes *C. albicans* has are β -galactosaminidase and L-proline aminopeptidase, those substrates (MNGL, 4-methylumbelliferyl-*N*-acetyl- β -D-galactosaminide for β -galactosaminidase; fluorogenic and PRO, L-proline *p*-nitroanilide for L-proline aminopeptidase; chromogenic) are infused on a cotton swab in this kit. After picking up a single colony with the swab, one can determine within approximately 1 hour whether the isolate is *Candida albicans*. The procedure is very simple: pick-up of one colony with a swab, incubation for 1 hour at 37°C with one drop of accompanying buffer, then visualization with a UV lamp (366 nm) and addition of a color

Corresponding author: Reiko Tanaka

Research Center for Pathogenic Fungi and Microbial
Toxicoses, Chiba University

1-8-1 Inohana, Chuo-ku, Chiba, 260-8673 Japan

Table 1. Specificity of RID Zyme CAS test

Organism	No. tested	MNGL	PRO
<i>Candida albicans</i> (serotypes A, B)	1136	+	+
<i>C. dubliniensis</i>	21	+	+
<i>C. stellatoidea</i>	5	+	-
<i>C. glabrata</i>	54	-	-
<i>C. guilliermondii</i>	15	-	+
<i>C. kefyr</i>	4	-	-
<i>C. krusei</i>	20	-	-
<i>C. parapsilosis</i>	138	-	+
<i>C. tropicalis</i>	107	+	-
<i>Cryptococcus neoformans</i> (serotypes A, B, C)	3	-	-
<i>Cr. neoformans</i> (serotypes D, AD)	2	+	-

MNGL, 4-methylumbelliferyl-*N*-acetyl- β -D-galactosaminide; PRO, L-proline *p*-nitroanilide.
+, positive; -, negative.

Table 2. Non-*albicans* *Candida* species tested

Species distinguishable from <i>C. albicans</i> by RID Zyme CAS test	
Clinically important species	
<i>glabrata</i> (CBS 138 ^T and 53 isolates)	
<i>guilliermondii</i> (ATCC 22995 ^T and 14 isolates)	
<i>kefyr</i> (ATCC 4135 ^T and 3 isolates)	
<i>krusei</i> (IFO 1395 ^T and 19 isolates)	
<i>parapsilosis</i> (IFM 5751 and 137 isolates)	
<i>stellatoidea</i> (ATCC 11006 ^T and 4 isolates)	
<i>tropicalis</i> (IFM 41420 and 106 isolates)	
Others	
<i>apicola</i> (ATCC 24616 ^T)	<i>mesenterica</i> (IFO 1123 ^T)
<i>boleticola</i> (CBS 6420 ^T)	<i>mogii</i> (IFO 0436 ^T)
<i>bombi</i> (ATCC 18811 ^T)	<i>molischiana</i> (IFO 10296 ^T)
<i>chilensis</i> (ATCC 22076 ^T)	<i>norvegensis</i> (CBS 1922 ^T)
<i>chiropterorum</i> (ATCC 22291 ^T)	<i>oleophila</i> (IFO 1021 ^T)
<i>cylindracea</i> (ATCC 14830 ^T)	<i>pelliculosa</i> (IFM 47124)
<i>famata</i> (IFM 47882)	<i>saitoana</i> (ATCC 36584 ^T)
<i>haemulonii</i> (ATCC 22991 ^T)	<i>santamariae</i> (IFO 1982 ^T)
<i>holmii</i> (IFO 1128 ^T)	<i>santamariae</i> var. <i>membranifaciens</i> (CBS 5838 ^T)
<i>inconspicua</i> (CBS 180 ^T)	<i>silvatica</i> (IFO 10311 ^T)
<i>intermedia</i> (ATCC 14439 ^T)	<i>sphaerica</i> (IFM 48789)
<i>lipolytica</i> (IFM 5475)	<i>torresii</i> (IFO 10421 ^T)
<i>lusitaniae</i> (IFM 49723)	<i>tsuchiyae</i> (IFO 10167 ^T)
<i>magnoliae</i> (CBS 166 ^T)	<i>utilis</i> (IFM 40125)
<i>melibiosica</i> (IFO 10238 ^T)	<i>viswanathii</i> (CBS 4024 ^T)
<i>melinii</i> (IFM 5473)	<i>zeylanoides</i> (IFO 1663 ^T)
Species indistinguishable from <i>C. albicans</i> by RID Zyme CAS test	
<i>kruisii</i> * (ATCC 24408 ^T)	<i>catenulata</i> (ATCC 10565 ^T)
<i>sake</i> * (NBRC 1354)	<i>dubliniensis</i> (CBS 7987 ^T and 20 isolates)
<i>savonica</i> * (IFO 10309 ^T)	<i>maltosa</i> (IFM 52017)
<i>suecica</i> * (IFO 10313 ^T)	<i>rugosa</i> (CBS 613 ^T)

*, No growth at 37°C; T, type strain.

development reagent (*p*-dimethylaminocinnamaldehyde). If the isolate is *C. albicans*, MNGL fluorescence is produced, and the PRO chromogen turns the swab purple (Fig. 1).

Results of the RID Zyme CAS test with nine clinically important *Candida* species and two *Cr. neoformans* species are shown in Table 1. All of the 1136 strains of *C. albicans* tested, including

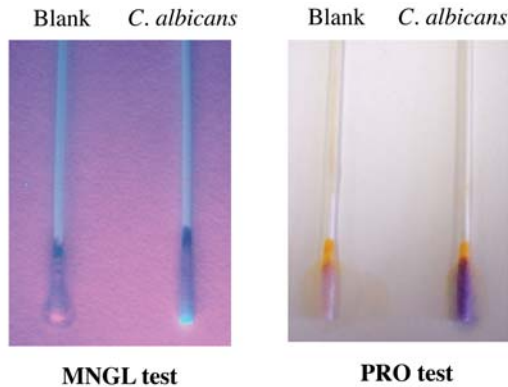


Fig. 1 Results of RID Zyme CAS test.

MNGL, 4-methylumbelliferyl-*N*-acetyl- β -D-galactosaminide;
PRO, L-proline *p*-nitroanilide.

serotype B strains, were positive in both tests (MNGL and PRO tests). A recently classified atypical *C. albicans* species, *C. dubliniensis*, was also positive in both tests. *C. stellatoidea*, about which the taxonomy difference with *C. albicans* has been argued^{4, 6-8)} and which now is recognized as a synonym of *C. albicans*, was only positive in the MNGL test. Clear distinction from *C. albicans* was obtained for *C. parapsilosis*, *C. guilliermondii*, *C. kefyr*, *C. glabrata*, *C. tropicalis*, *C. krusei*, and *Cr. neoformans*. With the exception of eight species (*C. catenulata*, *C. dubliniensis*, *C. kruisii*, *C. maltosa*, *C. rugosa*, *C. sake*, *C. savonica*, *C. suecica*), distinction from *C. albicans* was obtained with other non-*albicans* species (Table 2). Four of the eight indistinguishable species (*C. kruisii*, *C. sake*, *C. savonica*, *C. suecica*) showed no growth at 37°C, and four species (*C. catenulata*, *C. dubliniensis*, *C. maltosa*, *C. rugosa*) showed no distinction from *C. albicans* with the kit. However, *C. maltosa* has been reported as non-pathogenic for mice⁹⁾, and distinction of *C. catenulata* and *C. rugosa* from *C. albicans* can be made microscopically¹⁰⁾. Summarizing these data, the sensitivity of this kit was 100% and the specificity was 97.6%. On the other hand, Crist *et al.*¹¹⁾ and Heelan *et al.*¹²⁾ compared four methods (MUREX *C. albicans*, Albicans-Sure, BactiCard *Candida* and the germ tube test), when the operation time of BactiCard *Candida* and Albicans-Sure was being emphasized as 5 minutes. Although the operation time of the RIDzyme CAS test is 1 hour, it is superior to those two methods in that one colony is sufficient for the inoculation. The specificity of those four methods apparently was higher than the RIDzyme CAS test since *C. dubliniensis* was not taken into consideration. According to these results, the RID

Zyme CAS test kit is effective in the identification of *C. albicans* from clinical samples. Unfortunately, *C. dubliniensis*, which has been increasingly reported in recent years, was indistinguishable from *C. albicans*.

This study was performed as part of the Frontier Studies and International Networking of Genetic Resources in Pathogenic Fungi and Actinomycetes (FN-GRPF) through the Special Coordination Funds for Promoting Science and Technology from the Ministry of Education, Culture, Sports, Science and Technology of Japan (2003), and Grant-in-Aid for Scientific Research (C-16510176) from Japan Society for the Promotion of Science.

REFERENCES

- 1) Perry JL, Miller GR: Umbelliferyl-labeled galactosaminide as an aid in identification of *Candida albicans*. *J Clin Microbiol* **25**: 2424-2425, 1987.
- 2) Dalton MT, Haldane DJ, MacDonald J: Rapid identification of *Candida albicans* using 4-methylumbelliferyl *N*-acetyl- β -galactosaminide. *Diagn Microbiol Infect Dis* **12**: 521-523, 1989.
- 3) Perry JL, Miller GR, Carr DL: Rapid, colorimetric identification of *Candida albicans*. *J Clin Microbiol* **28**: 614-615, 1990.
- 4) McCullough MJ, Clemons KV, Stevens DA: Molecular and phenotypic characterization of genotypic *Candida albicans* subgroups and comparison with *Candida dubliniensis* and *Candida stellatoidea*. *J Clin Microbiol* **37**: 417-421, 1999.
- 5) Kanbe T, Horii T, Arishima T, Ozeki M, Kikuchi A: PCR-based identification of pathogenic *Candida* species using primer mixes specific to *Candida* DNA topoisomerase II genes. *Yeast* **19**: 973-989, 2002.
- 6) Biswas SK, Yokoyama K, Wang L, Nishimura K, Miyaji M: Typing of *Candida albicans* isolates by sequence analysis of the cytochrome *b* gene and differentiation from *Candida stellatoidea*. *J Clin Microbiol* **39**: 1600-1603, 2001.
- 7) Kwon-Chung KJ, Hicks JB, Lipke PN: Evidence that *Candida stellatoidea* type II is a mutant of *Candida albicans* that does not express sucrose-inhibitable α -glucosidase. *Infect Immun* **58**: 2804-2808, 1990.
- 8) Timmins EM, Howell SA, Alsberg BK, Noble WC, Goodacre R: Rapid differentiation of closely related *Candida* species and strains by pyrolysis-mass spectrometry and Fourier transform-infrared spectroscopy. *J Clin Microbiol* **36**: 367-374, 1998.
- 9) Holzschu DL, Chandler FW, Ajello L, Ahearn DG: Evaluation of industrial yeasts for pathogenicity. *Sabouraudia* **17**: 71-78, 1979.

- 10) Barnett L: Barnett JA, Payne RW, Yarrow D (ed.), *Yeasts: Characteristics and Identification*, 3rd ed. Cambridge University Press, Cambridge, UK, 2000.
- 11) Crist AE Jr, Dietz TJ, Kampschroer K: Comparison of the MUREX *C. albicans*, Albicans-Sure, and BactiCard Candida test kits with the germ tube test for presumptive identification of *Candida albicans*. *J Clin Microbiol* **34**: 2616-2618, 1996.
- 12) Heelan JS, Siliezar D, Coon K: Comparison of rapid testing methods for enzyme production with the germ tube method for presumptive identification of *Candida albicans*. *J Clin Microbiol* **34**: 2847-2849, 1996.